

## Isolation of Bacteria from Rhizosphere, Phylloplane and Caulosphere of Brinjal (*Solanum melongena* L.)

Jennifer Lolitha C.<sup>1</sup>, Manjula A. C.<sup>2</sup>, Prathibha K. Y.<sup>3</sup> and Keshamma E.<sup>4</sup>

<sup>1</sup>Associate Professor, Department of Botany, Maharani Cluster University, INDIA.

<sup>2</sup>Associate Professor, Department of Sericulture, Maharani Cluster University, INDIA.

<sup>3</sup>Associate Professor, Department of Botany, Maharani Cluster University, INDIA.

<sup>4</sup>Assistant Professor, Department of Biochemistry, Maharani Cluster University, INDIA.

<sup>4</sup>Corresponding Author: keshamma.blr76@gmail.com

### ABSTRACT

Rhizosphere, phylloplane and caulosphere is the region where a complex community of microbes, mainly bacteria and fungi are present. The microbe-plant interaction in these regions can be beneficial, neutral, variable, or deleterious for plant growth. The bacteria that exert beneficial effects on plant development are termed plant growth promoting bacteria. To isolate the bacteria from rhizosphere, phylloplane and caulosphere of brinjal (*Solanum melongena* L.). The seeds of 16 cultivars of brinjal (*Solanum melongena* L.) viz., Arka keshav, Arka shirish, Arka kusumaker, and IIHR accession numbers 389,386,387,377 Tc, BB44, 391, 433, 434, 427, 447, 448, 476 and 487 that were used in the initial screening experiment were obtained from the Department of Vegetable crops, IIHR, Hesaraghatta, Bangalore. Brinjal (*Solanum melongena* L.) plants of different varieties were collected from seven locations around Bangalore viz., Hesaraghatta, Yelahanka, Kengeri, Madi vala, Hebbal, Tirumalapura and Attibele were also screened for the presence of associative bacteria. Associative microorganisms isolated from the rhizosphere, phylloplane and shoot regions of brinjal (*Solanum melongena* L.), revealed the presence of three morphologically different colonies. 80% of 16 cultivars of the brinjal (*Solanum melongena* L.) screened showed the presence of associative bacterial colonies. In this study diazotrophic BBI were obtained from the rhizoplane, phylloplane and stem of 16 cultivars of brinjal (*Solanum melongena* L.) that were screened. The dominant pearl-colored colonies isolated from all varieties of brinjal plants that were screened was identified and showed maximum nitrogen fixing ability compared with that of the other colonies. The phylloplane of brinjal (*Solanum melongena* L.) plants from seven different locations around Bangalore showed the presence of the dominant pearl-colored colonies. Moderate growth of bacteria was observed in root, stem and leaf bits sterilized up to 35 minutes. Even on surface sterilized roots which were homogenized and inoculated on growth media, dense growth of bacteria was observed there by establishing the presence of bacteria inside the root system. For the first time the presence of growth promoting bacteria on the rhizosphere and endorhizosphere of brinjal (*Solanum Melongena* L.) cultivars was established.

**Keywords-** Bacteria, Rhizosphere, Phylloplane, Caulosphere, *Solanum melongena* L.

### I. INTRODUCTION

Although symbiotic Nitrogen fixation especially legume-rhizobium system has been proved to be the best form of biological Nitrogen fixation, associative nitrogen fixation cannot be ignored. Nitrogen fixation on the rhizoplane, phylloplane and stem have been attributed to the presence of diazotrophic bacteria associated with the roots, stem and leaves of plants [1]. Associative bacteria have been isolated from the rhizoplane, phylloplane and stem of many non-leguminous plants. Many studies have dealt with isolation of associative microorganisms from the roots of cereals, vegetable and fruit crops such as sweet potato [2], arecanut, banana, coconut, cashew, citrus, custard apple, grape, guava, jackfruit, litchi, mango, papaya, pomegranate- ate, phalsa, pepper, and strawberry [3], *Spartina altemifolia* [4], sugarcane [5], barley [6], wheat, maize, sorghum, millet and rice [7-12]. Root Smiace colonization by *Azospirillum* species in tomato, pepper, cotton and cucumis sativa under normal condition were studied by [13][14]. The bacteria were found efficiently colonizing the root elongation and root hair zones of the above plants. These isolated bacteria had growth promoting properties [15][16]. Isolated *Azospirillum* from the rhizosphere of tomatoes, white potatoes and corn [17]. Reported the presence of a new species of *Azospirillum bangaloreense* colonizing the endorhizosphere of tomato. Studies on many graminaceous plants showed the presence of associated bacteria from the interiors of various plant parts [18-21] did similar studies relating to isolation of nitrogen fixing bacteria associated with the roots and rhizosphere of maize and other cereals and grasses [22]. Isolated a root colonizing bacterium, which was characterized as *Pseudomonas* strain from roots of potato. Nitrogen fixing organisms were isolated from the roots of many non- leguminous crops such as *Spicacia oleracea*, *Brassica chinensis* and *Brassica rapa* by [23]. The isolated associated bacteria in many of these studies was identified as *Azospirillum* [17][19][24-28] observed spatial distribution of associated microorganisms identified as *Azospirillum* in rhizosphere of barley plants [29]. Isolated two nitrogen-fixing bacteria from

the rhizosphere of mangrove trees, which were characterized as [2] characterized a nitrogen-fixing bacteria associated with the roots of sweet potato as *Azospirillum*. In the present study both sterile and unsterile root, leaf and stem bits of 16 cultivars of brinjal (*Solanum melongena* L.) plants were used for the initial screening of associative bacteria. The dominant colonies of bacteria present in the rhizoplane, phylloplane and stem were isolated.

## II. METEIRALS AND MATHODS

The seeds of 16 cultivars of brinjal (*Solanum melongena* L.) viz., *Arka keshav*, *Arka shirish*, *Arka kusumaker*, and IIHR accession numbers 389,386,387,377 Tc, BB44, 391, 433, 434, 427, 447, 448, 476 and 487 that were used in the initial screening experiment were obtained from the Department of Vegetable crops, IIHR, Hessaraghatta, Bangalore. Brinjal (*Solanum melongena* L.) plants of different varieties were collected from seven locations around Bangalore viz., Hessaraghatta, Yelahanka, Kengeri, Madi vala, Hebbal, Tirumalapura and Attibele were also screened for the presence of associative bacteria. The standard laboratory chemicals were used and Bacteriological media used were from Himedia.

The procedure given by Patriquin and [1] was followed to isolate the associative bacteria from the root, stem and leaf of brinjal. The cultivars were grown in '10 x 10cm clay pots filled with sandy loam soil and topped with farm yard manure (FYM) for a period of 2 months. Sixteen brinjal cultivars were screened for the presence of associative microorganisms. The plants were carefully removed from the pots and the roots system was washed with sterile distilled water and was cut into bits measuring 0.5 cms. The bits were divided into two sets, one set was surface sterilized by immersing in 0.1% mercuric chloride for 2-3 minutes followed by washing with distilled water. Both control and surface sterile root bits were inoculated individually into test tubes containing 5 ml of sterile solid nitrogen free Burk's media [8]. The inoculated test tubes were incubated at 35° C for 48 hours. The bacterial growth from the sterile roots was isolated and sub cultured in nitrogen free Burk's media. They were multiplied in TYMB media

[30]. Pure cultures were maintained in stab cultures containing nitrogen free Burk's media.

Leaves were separated from the stem, washed in running water and cut into 0.2 cms long strips followed by washing with sterile distilled water thrice. One set was surface sterilized by immersing in 0.1 % mercuric chloride for 2-3 minutes. Both control and surface sterilized leaf bits were inoculated into test tubes containing 5ml of semi solid nitrogen free Burk's media [8], and incubated at 37°C for 48 hours. The bacteria were isolated from sterile leaf tissues and subcultured on nitrogen free Burk's media. Pure cultures were maintained in stab cultures, containing nitrogen free Burk's media. Associated bacteria from the stem of the above mentioned 16 cultivars of brinjal (*Solanum melongena* L.) were isolated using the same procedure that was followed for the isolation of associative bacteria from the rhizosphere. Pure cultures were maintained in stab cultures containing nitrogen free Burk's media.

## III. RESULT

Associative microorganisms isolated from the rhizosphere, phylloplane and shoot regions of brinjal (*Solanum melongena* L.), revealed the presence of three morphologically different colonies. (Table-1) 80% of 16 cultivars of the brinjal (*Solanum melongena* L.) screened showed the presence of associative bacterial colonies. (Table-2) In this study diazotrophic brinjal bacterial isolate (BBI) were obtained from the rhizoplane, phylloplane and stem of 16 cultivars of brinjal (*Solanum melongena* L.) that were screened. The dominant pearl-colored colonies isolated from all varieties of brinjal plants that were screened was identified and chosen for the present study as it showed maximum nitrogen fixing ability compared with that of the other colonies. The phylloplane of brinjal (*Solanum melongena* L.) plants from seven different locations around Bangalore showed the presence of the dominant pearl-colored colonies. Moderate growth of bacteria was observed in root, stem and leaf bits sterilized up to 35 minutes (Plate-1). Even on surface sterilized roots which were homogenized and inoculated on growth media, dense growth of bacteria was recorded there by establishing the presence of bacteria inside the root system. (Table-3)

**Table 1: Isolation of associative bacteria from sterilized and unsterilized rhizoplane, phylloplane and stem of 16 IIHR varieties of brinjal (*Solanum melongena* L.).**

IIHR Acc. Number	Rhizoplane		Phylloplane		Stem	
	Unsterilised	Sterilised	Unsterilised	Sterilised	Unsterilised	Sterilised
389	+	• + •	+ •	+	+ •	+ •
386	+	+	+ •	+	+ •	+
387	+ •	+	+ •	+	+ -	+ -
377 TC	+ -	+	+ •	+ •	+	+
<i>Arka keshav</i>	+	• + •	+ •	+	+ •	+
<i>Arka shirish</i>	+ •	+ •	+ •	+	+ •	+ •

<i>Arka kusumakar</i>	♦ -	+ ♦	+ ♦	+ ♦	+ ♦	+ ♦
8844	+ ♦ -	+ ♦	+ ♦	+ ♦	+ ♦	+ ♦
391	+ -	+	+	+	+ ♦ -	+
433	+	+	+	+	+ -	+
434	+ ♦	+ ♦	+ ♦	+	+ -	+ ♦
427	+ ♦ -	+ ♦	+ ♦	+	+ ♦	+ ♦
447	♦ -	♦	+	+	+ ♦	+ ♦
448	+	+	+	+ ♦	+ ♦	+
476	+ ♦	+	+	+	+	+
487	+ ♦	+ ♦	+ ♦	+ ♦	+ ♦	+ ♦

- + Indicates pearl coloured
- ♦ - Indicates cream-coloured colonies
- Indicates cream-coloured colonies

Table 2: Bacterial growth on brinjal root, stem and leaf sterilized for different time periods

Sterilization (min)	Bacterial growth on		
	Stem	Root	Leaf
2	++	++++	+++
5	++	++++	+++
10	+	+++	++
15	+	++	+
20	-	+	
30			
60			

++++ very high, +++ high, ++ moderate, + low, - nil



Plate 1: Phyllosphere bacteria in brinjal CV. Arka Keshav

Table 3: Isolation of Associative Bacteria from sterilized rhizoplane, phylloplane and stem of brinjal (*Solanum melongena* L.) plants collected from different locations around Bangalore.

Locations	Bacterial growth on		
	Stem	Root	Leaf
Attibele	+	+	+
Hebbal	+	+	+

Hessaraghatta	+	+	+
Kengeri	+	+	+
Madivala	+	+	+
Tirumalapura	+	+	+
Yalahanka	+	+	+

+ - indicates pearl colored

#### IV. DISCUSSION

The presence of growth promoting bacteria on the rhizosphere and endorhizosphere of brinjal (*Solanum Melongena* L.) cultivars was established in the present study for the first time. Though [31][32] in a preliminary study isolated acetylene-reducing bacteria from the rhizosphere of eggplant they did not unequivocally prove their presence in the endorhizosphere or their growth promoting ability. In this study the dominant colonies of associative bacteria were recorded on the surface of stem, root and leaves of brinjal. Studies revealed that these bacteria developed a symbiotic relationship with the root system of the plant colonizing the intercellular and intracellular spaces of the cortex and the stem and the leaf through the conducting tissues. Growth promoting bacteria have been isolated in vegetables like tomato, cabbage, spinach, winged bean, capsicum and sweet potato [13][33]. observed aggregates of bacteria on the surface and endosphere of root hair, root cap and elongation zones of tomato using scanning rhizobacteria. Similar isolations have been reported from roots of cereals, grasses and plantation crops [8].

#### V. CONCLUSION

The study isolated bacteria which possess several plant growths promoting traits. This reveals the potential of these strains for biofertilizer application and commercial use as biocontrol agents in the field. Thus, these isolates can perform close to its optimum potential. Future studies concerning commercial and field applications of integrated stable bio-formulations as effective biocontrol are needed.

#### REFERENCES

[1] Dobereiner, J. (1980). Forage grasses and grain crops. Methods for evaluating biological nitrogen fixation, 535-536.  
 [2] Hill, W. A., Bacon-Hill, P., Crossman, S. M., & Stevens, C. (1983). Characterization of N<sub>2</sub>-fixing bacteria associated with sweet potato roots. Canadian journal of microbiology, 29(8), 860-862.  
 [3] Nair, S. K., & Rao, N. S. (1977). Microbiology of the root region of coconut and cacao under mixed cropping. Plant and Soil, 46(3), 511-519.

[4] McClung, C. R., van Berkum, P., Davis, R. E., & Sloger, C. (1983). Enumeration and localization of N<sub>2</sub>-fixing bacteria associated with roots of *Spartina alterniflora* Loisel. Applied and Environmental Microbiology, 45(6), 1914-1920.  
 [5] Graciolli, L. A. (1981). Microorganisms in the phyllosphere and rhizosphere of sugarcane. Associative N<sub>2</sub> Fixation, 91-101.  
 [6] Pohlman, A. A., & McColl, J. G. (1982). Nitrogen fixation in the rhizosphere and rhizoplane of barley. Plant and Soil, 69(3), 341-352.  
 [7] Dart, P. J., & Wani, S. P. (1982). Non-symbiotic nitrogen fixation and soil fertility.  
 [8] Rao, N. S. (1983). Nitrogen-fixing bacteria associated with plantation and orchard plants. Canadian journal of microbiology, 29(8), 863-866.  
 [9] Klucas, R. V., Pedersen, W., Shearman, R. C., & Wood, L. V. (1981). Nitrogen fixation associated with winter wheat, sorghum, and Kentucky bluegrass [*Triticum aestivum*, *sorghum bicolor*, *Poa pratensis*].  
 [10] Kumari, M. L., Kavimandan, S. K., & Subba Rao, N. S. (1970). Occurrence of nitrogen fixing Spirillum in roots of rice, sorghum, maize & other plants. Indian Journal of Experimental Biology, 14(5), 638-639.  
 [11] Patriquin, D. G., & Döbereiner, J. (1978). Light microscopy observations of tetrazolium-reducing bacteria in the endorhizosphere of maize and other grasses in Brazil. Canadian journal of microbiology, 24(6), 734-742.  
 [12] Patriquin, D. G., Döbereiner, J., & Jain, D. K. (1983). Sites and processes of association between diazotrophs and grasses. Canadian Journal of Microbiology, 29(8), 900-915.  
 [13] Bashan, Y., & Holguin, G. (1986). Anchoring of *Azospirillum brasilense* to hydrophobic polystyrene and wheat roots. Microbiology, 139(2), 379-385.  
 [14] Bashan, Y., Levany, H., & Whitmoyer, R. E. (1991). Root surface colonization of non-cereal crop plants by pleomorphic *Azospirillum brasilense* Cd. Microbiology, 137(1), 187-196.  
 [15] Quimio, A. J., & Coroza, A. G. (1986). Plant growth promoting activity of rhizobacteria on tomato, white potato and corn [Philippines]. Supplement No. 1.  
 [16] Quimio AJ, Cadapan ET. Field performance of selected plant growth promoting rhizobacteria (PGPR) on corn, sorghum and cowpea. Philippine Journal of Crop Science. 1988.  
 [17] Sukhada M. (1987). Field response of tomato (*Lycopersicon esculentum* Mill 'Pusa Ruby') to

inoculation with a VA mycorrhizal fungus *Glomus fasciculatum* and with *Azotobacter vinelandii*. Plant and soil, 98(2), 295-297.

[18] Bashan, Y. (1983). Enhancement of wheat root colonization and plant development by *Azospirillum brasilense* Cd. following temporary depression of rhizosphere microflora. Applied and Environmental Microbiology, 51(5), 1067-1071.

[19] Bashan, Y., Harrison, S. K., & Whitmoyer, R. E. (1990). Enhanced growth of wheat and soybean plants inoculated with *Azospirillum brasilense* is not necessarily due to general enhancement of mineral uptake. Applied and Environmental Microbiology, 56(3), 769-775.

[20] Boddey, R. M., & Dobreiner, J. (1988). Nitrogen fixation associated with grasses and cereals: recent results and perspectives for future research. Plant and soil, 108(1), 53-65.

[21] Lalande, R., Bissonnette, N., Coutlee, D. and Antoun, H. (1989). Plant and Soil (Netherlands);117 (2): 207-218.

[22] Brand, I., Lugtenberg, B. J. J., Glandorf, D. C. M., Bakker, P. A. H. M., Schippers, B., & De Weger, L. A. (1991). Isolation and characterization of a superior potato root-colonizing *Pseudomonas* strain. Bulletin OILB SROP (France).

[23] Ahn, S. B., Gamo, H., & Yuk, C. S. (1990). Isolation of N<sub>2</sub>-fixing Microorganism from the Root of Non-leguminous Crops. Korean Journal of Soil Science and Fertilizer, 23(1), 67-72.

[24] Tyler, M. E., Milam, J. R., Smith, R. L., Schank, S. C., & Zuberer, D. A. (1979). Isolation of *Azospirillum* from diverse geographic regions. Canadian Journal of Microbiology, 25(6), 693-697.

[25] Malik, K. A., & Bilal, R. (1989). Survival and colonization of inoculated bacteria in kallar grass rhizosphere and quantification of N<sub>2</sub>-fixation. In Nitrogen Fixation with Non-Legumes (pp. 301-310). Springer, Dordrecht.

[26] Bilal, R., Rasul, G., Arshad, M., & Malik, K. A. (1993). Attachment, colonization and proliferation of *Azospirillum brasilense* and *Enterobacter* spp. on root surface of grasses. World Journal of Microbiology and Biotechnology, 9(1), 63-69.

[27] Agarwala-Dutt, R., Tilak, K. V. B. R., & Rana, J. P. S. (1991). Isolation of *Azospirillum* from the interior of various parts of some graminaceous plants. Zentralblatt für Mikrobiologie, 146(3), 217-219.

[28] Lukin, S., Kozhevin, P., & Zviagintsev, D. (1990). Spatial distribution of *Azospirillum brasilense* cells in the rhizosphere of barley plants. Mikrobiologiya (Moskva, 1932), (6), 1090-1093.

[29] Holguin, G., Guzman, M. A., & Bashan, Y. (1992). Two new nitrogen-fixing bacteria from the rhizosphere of mangrove trees: Their isolation, identification and in vitro interaction with rhizosphere *Staphylococcus* sp. FEMS Microbiology Letters, 101(3), 207-216.

[30] Dalton, H. (1980). The cultivation of diazotrophic microorganisms. The cultivation of diazotrophic microorganisms., 13-64.

[31] Gamo, T. (1990). *Azospirillum* spp. from crop roots: a promoter of plant growth. JARQ. Japan Agricultural Research Quarterly, 24(4), 253-259.

[32] Gamo, T., & Ahn, S. B. (1991). Growth-promoting *Azospirillum* spp. isolated from the roots of several non-gramineous crops in Japan. Soil Science and Plant Nutrition, 37(3), 455-461.

[33] Bashan, Y., Singh, M., & Levanony, H. (1989). Contribution of *Azospirillum brasilense* Cd to growth of tomato seedlings is not through nitrogen fixation. Canadian Journal of Botany, 67(8), 2429-2434.